AD	1	
æ		

Award Number: DAMD17-01-1-0352

TITLE: Neural Protein Synuclein Gamma (SNCG) in Breast Cancer

Progression

PRINCIPAL INVESTIGATOR: Yangfu Jiang, M.D., Ph.D.

CONTRACTING ORGANIZATION: Long Island Jewish Medical Center

Lake Success, New York 11042

REPORT DATE: July 2003

TYPE OF REPORT: Annual Summary

PREPARED FOR: U.S. Army Medical Research and Materiel Command

Fort Detrick, Maryland 21702-5012

DISTRIBUTION STATEMENT: Approved for Public Release;

Distribution Unlimited

The views, opinions and/or findings contained in this report are those of the author(s) and should not be construed as an official Department of the Army position, policy or decision unless so designated by other documentation.

REPORT DOCUMENTATION PAGE

Form Approved OMB No. 074-0188

Public reporting burden for this collection of information is estimated to average 1 hour per response, including the time for reviewing instructions, searching existing data sources, gathering and maintaining the data needed, and completing and reviewing this collection of information. Send comments regarding this burden estimate or any other aspect of this collection of information, including suggestions for reducing this burden to Washington Headquarters Services, Directorate for Information Operations and Reports, 1215 Jefferson Davis Highway, Suite 1204, Arlington, VA 22202-4302, and to the Office of Management and Budget, Paperwork Reduction Project (0704-0188). Washington, DC 20503

1. AGENCY USE ONLY (Leave blank)	2. REPORT DATE July 2003	3. REPORT TYPE AND DATES COVERED Annual Summary (1 Jul 2002 - 30 Jun 2003)
4. TITLE AND SUBTITLE Neural Protein Synuclein Progression	Gamma (SNCG) in Brea	5. FUNDING NUMBERS DAMD17-01-1-0352

6. AUTHOR(S)

Yangfu Jiang, M.D., Ph.D.

7. PERFORMING ORGANIZATION NAME(S) AND ADDRESS(ES)

Long Island Jewish Medical Center Lake Success, New York 11042

8. PERFORMING ORGANIZATION REPORT NUMBER

E-Mail: jyf11040@yahoo.com

9. SPONSORING / MONITORING AGENCY NAME(S) AND ADDRESS(ES)

U.S. Army Medical Research and Materiel Command Fort Detrick, Maryland 21702-5012

10. SPONSORING / MONITORING AGENCY REPORT NUMBER

11. SUPPLEMENTARY NOTES

12a. DISTRIBUTION / AVAILABILITY STATEMENT

Approved for Public Release; Distribution Unlimited

12b. DISTRIBUTION CODE

13. ABSTRACT (Maximum 200 Words)

Synucleins are emerging as a central player in the fundamental neural processes and in the formation of pathologically insoluble deposits characteristic of Alzheimer's (AD) and Parkinson's (PD) diseases. However, the normal cellular function of this highly conserved synuclein family remains largely unknown. Using differential cDNA sequencing approach, we first identified a breast cancer specific gene, BCSG1, which was expressed abundantly in metastatic breast cancer cDNA library but scarcely in normal breast cDNA library. Interestingly, BCSG1 revealed no homology to any other known growth factors or oncogenes; rather, BCSG1 revealed extensive sequence homology to neurotic proteins of α synuclein and β synuclein, and thus was also named as γ Synuclein (SNCG). SNCG expression is highly associated with breast cancer and ovarian cancer progression. In addition, overexpression of SNCG in breast cancer cells significantly stimulated cell growth in vitro and tumor metastasis in vivo. However, the molecular targets of SNCG aberrant expression for breast cancer have not been identified. For the first time, we report a chaperone-like activity of SNCG in stimulating the transcriptional activity of estrogen receptor- α (ER- α) in breast cancer cells. Consistent with the stimulation of ER- α , SNCG stimulated the ligand-dependent cell proliferation, suppression of endogenous SNCG expression significantly inhibited cell growth in response to estrogen. The stimulatory effect of SNCG on ER α -regulated gene expression and cell growth can be effectively inhibited by antiestrogens. Demonstration of the stimulation of ER- α signaling as one of the cellular functions of SNCG will have a great impact on the biology of steroid receptors and the pathological role of SNCG on hormone-responsive tumors including breast, ovary, and prostate.

14. SUBJECT TERMS							
	15. NUMBER OF PAGES						
No Subject Terms Provi	10						
	1						
			16. PRICE CODE				
	70. PRICE CODE						
17. SECURITY CLASSIFICATION	18. SECURITY CLASSIFICATION	40 0504DITY 01 4004TIC 1-1014	-				
		19. SECURITY CLASSIFICATION	20. LIMITATION OF ABSTRACT				
OF REPORT	OF THIS PAGE	OF ABSTRACT					
Unclassified	Unclassified	Unclassified					
	00143511164	Unclassified	Unlimited				

NSN 7540-01-280-5500

Standard Form 298 (Rev. 2-89) Prescribed by ANSI Std. Z39-18 298-102

Table of Contents

Cover	1
SF 298	2
Table of Contents	3
Introduction	4
Research Report	. 4-8
Key Research Accomplishments	8
Conclusions	8-9
References	9-10

A. INTRODUCTION

- A-1. Identification of genes differentially expressed in breast cancer versus normal breast. We undertook a search, using the differential cDNA sequencing approach as we previously described (1-3), for isolation of differentially expressed genes in the cDNA libraries from normal breast and breast carcinoma. Of many putative differentially expressed genes, a breast cancer specific gene, BCSG1, which was (a) highly expressed in mammary gland relative to other organs and was (b) high abundance in a breast cancer cDNA library but scarcely in a normal breast cDNA library, was identified as a putative breast cancer marker (1). We demonstrated a stage-specific BCSG1 expression as follows: BCSG1 was undetectable in normal or benign breast lesions, showed partial expression in ductal carcinoma in situ, but was expressed at an extremely high level in advanced infiltrating breast cancer (1). Overexpression of BCSG1 was also demonstrated in ovarian cancer (4).
- A-2. Neural protein synuclein. Interestingly, BCSG1 revealed no homology to any other known growth factors or oncogenes; rather, BCSG1 revealed extensive sequence homology to neurotic protein synuclein, having 54% and 56% sequence identity with α synuclein (SNCA) and β synuclein (SNCB), respectively. Subsequent to the isolation of BCSG1, synuclein γ (5) and persyn (6) were independently cloned from a brain genomic library and a brain cDNA library. In fact, BCSG1, SNCG, and persyn appear to be the same protein. Thus, the previously identified BCSG1, which is also highly expressed in brain (1), has been renamed as SNCG as the third member of synuclen family (7). Synucleins has been specifically implicated in neurodegenerative diseases such as Alzheimer's disease (AD) and Parkinson's disease (PD). Mutations in SNCA is genetically linked to several independent familial cases of PD (8). More importantly, wild type of SNCA is the major component of Lewy bodies in sporadic PD (9-10). SNCA peptide known as non-amyloid component of plaques has been implicated in amyloidogenesis in AD (11-12). SNCB and SNCG have also been recognized to play a role in the pathogenesis of PD and Lewy bodies cases (13-14).
- A-3. Expression of BCSG1/SNCG in breast and ovary cancer. Although synucleins are highly expressed in neuronal cells and are abundant in presynaptic terminals, synucleins have also been implicated in nonneural diseases particularly in the hormone responsive cancers of breast and ovary (1,4-5,15-17). Being identified as a breast cancer specific gene, SNCG mRNA was detected in neoplastic breast epithelial cells but not in normal mammary epithelial cells (1). While the expression of SNCG in normal breast is nondetectable (0 out of 7 normal breast specimens), 43% of stage II/III breast carcinomas (6 of 14) and 73% of stage IV breast carcinomas (11 of 15) expressed SNCG, respectively (Fig. 1). Western analysis to examine SNCG protein expression in human breast tissues showed a similar pattern in that it was not detected in normal breast tissues and stage I/II ductal breast carcinomas, but was detected in 70% of Stage III/IV ductal breast carcinomas (12). Ninkina et al were also able to confirm by using Northern and Western blotting that some breast tumors and breast tumor cell lines expressed SNCG, whereas normal breast tissue did not (16). In addition to the link to breast cancer progression, it has also been found that SNCG is involved in ovarian cancer. Following our identification of BCSG1, Lavedan et al first suggested that BCSG1/SNCG may be abnormally expressed in ovarian tumors as well as in breast tumors, based on the discovery of some SNCG ESTs in the libraries derived from an ovarian tumor (5). This suggestion was further confirmed by Western and immunohistochemical analyses (17). While synucleins (α , β , and γ) expression was not detectable by immunohistochemistry in normal ovarian epithelium, 87% (39 of 45) of ovarian carcinomas were found to express either SNCG or SNCB, and 42% (19 of 45) expressed all 3 synucleins (α , β , and γ) simultaneously.

B. RESEARCH REPORT

The original aims of establishing the MMTV/SNCG transgenic mice is underway. However, we are behind the schedule due to the virus infection in our small animal facility beginning last summer. In order to burn out the virus, the institution has imposed a 5-month no breeding period. We are currently in the screening process. In addition, we have entered the new research direction and got some exciting new data.

B-1. Specific expression of SNCG in advanced breast cancer. Previously, our *in situ* hybridization analysis has demonstrated an association between SNCG expression and breast cancer progression (1). To further evaluate the clinical relevance of SNCG expression to breast cancer progression in a large set of clinical specimens, we performed a semi-quantitative RT-PCR analysis on 79 clinical breast specimens including 12 normal or benign lesions, 2 DCIS, 41 stage I/II breast cancers, and 24 stage III/IV breast cancers. As shown in **Fig. 1** and summarized in **Table 1**, while no SNCG mRNA was detectable in 12 benign breast specimens, SNCG was expressed in most of advanced infiltrating breast cancers. The expression of SNCG mRNA was detectable in 26 of 67 breast cancer cases (38.8%). Among the 26 of SNCG positive cases, 19 cases were detected in the cancer with stage III/IV, only 6 cases were detected in the cancer with stage I/II, one case was detected in DCIS. There was no detectable SNCG mRNA in 3 cases with tumor size < 2 cm. Therefore, SNCG expression is a stage-specific for breast cancer with no detectable expression in benign lesion, 15% (6/41) expression in stage I/II breast carcinomas, and 79% (19/24) expression in stage III/IV breast carcinomas.

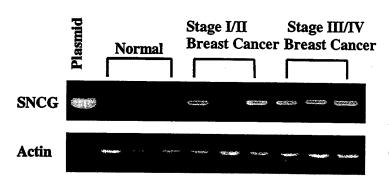


Fig. 1. Expression of SNCG in human breast tissue. Total RNA was isolated from frozen human breast specimens. RT-PCR analysis of SNCG expression was conducted using primers within SNCG cording sequence as described in Materials and Methods. The 384-bp PCR product is a specific indication of the presence of SNCG. The integrity and the loading control of the RNA samples were ascertained by actin expression

with a set of primers for 314-bp β -actin. A representative RT-PCR analysis was shown here with 3 benign breast lesions, 3 stage I/II breast cancer cases, and 3 stage III/IV breast cancer cases. The SNCG cDNA plasmid was used as a positive control.

Stage	Benign (n=12)	Stage I/II (n=41)	Stage III/IV (n=24)
Expression	0 (0%)	6 (15%)	19 (79%)

Table 1. Stage-specific expression of SNCG on breast cancer. Expression of SNCG in total 79 clinical breast specimens were investigated by 4 RT-PCR analyses. For each round of reaction, we used SNCG c-DNA plasmid as a template for the positive control. The integrity and the loading control of the RNA samples were ascertained by actin expression. Negative cases were confirmed with at least two independent experiments. These patients were randomly selected with the mean age of 58.2 years old, including 67 breast cancers, 7 breast hyperplasias, and 5 fibroadenomas.

B-2. Overexpression of SNCG stimulated transcriptional activity of ER- α . Estrogen response is mediated by two closely related members of the nuclear receptor family of transcription factors, ER- α and ER- β (18-19). Since ER- α is the major estrogen receptor in mammary epithelia, we measured the effect of SNCG on modulating the transcriptional activity of ER- α in human breast cancer cells. We first selected ER α -positive and SNCG-negative MCF-7 cells as recipient for SNCG transfection (Fig. 2A & B). MCF-7 cells were transiently transfected with either the pCI-SNCG expression plasmid or control pCI-neo plasmid. Transfection of SNCG gene into the SNCG-negative MCF-7 cells did not affect ER- α expression under the conditions both with and without E2 (Fig. 2A). In the absence of E2, the basal levels of ER- α on control and SNCG transfected cells are same. Although treatment of the control cells with E2 resulted in a significant decrease in ER- α level, overexpression of SNCG did not affect E2-mediated degradation of ER- α . Transfection of SNCG significantly stimulated E2-mediated activation of ER- α (Fig. 2B). Treatment of wild-type and SNCG transfected MCF-7 cells with E2 resulted in a significantly differential increase in estrogen-

responsive reporter ERE4-Luciferase (ERE4-Luc) activity relative to basal levels in untreated cells. Overexpression of SNCG gene in MCF-7 cells increased E2-stimulated reporter activity 3.2-fold over the SNCG-negative control cells. The SNCG-stimulated transcriptional activity of ER- α was ligand-dependent, because SNCG had no significant effect on the transcriptional activity of ER- α in the absence of E2.

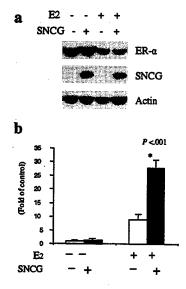


Fig. 2. SNCG stimulated ER-α transcriptional activity in MCF-7 human breast cancer cells. Cells were first transiently transfected with pCI-SNCG or the control vector pCI-neo. The transfected cells were selected with G418 and then transfected with pERE4-Luc as well as control reporter pRL-SV40-Luc. After transfection, cells were cultured in the ligand-free medium for 4 days as described in the Conditioned Cell Culture of Experimental Procedures, treated with or without 1 nM E2 for 24 hours before the promoter activities were determined by measuring the dual luciferase activity (A). Western analysis of ER-α and SNCG in MCF-7 cells transfected with pCI-SNCG or the control vector pCI-neo. Expression of SNCG did not affect the ER-α expression in the conditions both with and without 24-h E2 treatment. SNCG stimulated ER-α signaling in MCF-7 cells (B). The ERE reporter luciferase activity was normalized against the control renilla luciferase activity to correct for transfection efficiency. All values were presented as the fold induction over the control luciferase activity in the non-treated SNCG-negative cells, which

was taken as 1. The numbers represent means \pm SD of three cultures.

Consistent with the increased transcriptional activity of ER-α, SNCG also stimulated E2-regulated genes in MCF-7 cells (Fig. 3). While SNCG had no effect on the transcription of Cathepsin D (Cat-D), PS2, and TGF-α in the absence of E2, transcription of Cat-D, PS2, and TGF-α were increased 3.9-fold, 3.2-fold, and 4.2-fold in SNCG transfected cells vs. control cells in the presence of E2, respectively (Fig. 2A). To evaluate the effect of anti-estrogen on SNCG-stimulated ER-α-regulated genes, we treated the cells with an antiestrogen ICI 182,780 (ICI). As demonstrated in Fig. 2B, the basal levels of PR were very weak in both SNCG-transfected and control cells, but were increased significantly by E2 treatment. Treatment of the cells with E2 stimulated a 3.5-fold PR protein expression in SNCG transfected cells compared with control cells. Although ICI slightly stimulated basal levels of PR, treatment of the SNCG-transfected MCF-7 cells with ICI significantly blocked E2-stimulated PR expression, indicating that SNCG-stimulated gene expression in E2 treated cells is mediated by ER-α.

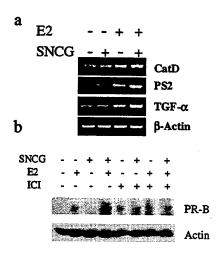


Fig. 3. SNCG stimulated estrogen-regulated gene transcription in MCF-7 cells. Cells were transiently transfected with pCI-SNCG or the control vector pCI-neo. After G418 selection, cells were cultured in the ligand-free medium for 4 days as described in the Conditioned Cell Culture of Experimental Procedures. A. RT-PCR. Cells were treated with or without 1 nM of E2 for 8 hours before the isolation of total RNA. Expressions of mRNA of Cat-D, PS2, and TGF-α were studied in SNCG transiently transfected cells vs. control cells by RT-PCR analyses. A an 842-bp product of Cat-D, a 336-bp product of PS2, and a 240-bp product of TGF-α, were amplified by RT-PCR and normalized with actin. B. Inhibition of SNCG-stimulated PR protein expression by antiestrogen ICI. Cells were treated with or without 1 nM of E2 and 1 μM of ICI for 32 h. Total proteins were isolated, normalized, and

subjected to Western analysis using anti-PR antibody.

B-3. Reduced levels of SNCG compromised transcriptional activity of ER-α. The effect of SNCG expression on

ER-α transactivation was further demonstrated by inhibiting endogenous SNCG expression with SNCG antisense mRNA in T47D cells that express high levels of SNCG (8). Stable transfection of the SNCG antisense construct into T47D cells significantly reduced SNCG expression to 25% of that in control T47D cells (Fig. 4A). While E2 significantly stimulated the reporter activity in the control T47D cells, inhibition of SNCG expression reduced E2-responsive luciferase activity in two stable antisense-transfected T47D cell lines, AS-1 and AS-3 cells, to 21% and 13% of that in control T47D cells, respectively (Fig. 4B). Treatment of T47D cells with E2 resulted in a 25-fold increase over the non-treated cells. However, only 3.3- and 5.2-fold increase was observed in the AS-3 and AS-1 cells, respectively. Taken together, the increased estrogen-responsive reporter activity in SNCG transfected MCF-7 and MDA-MB-435 cells as well as the increased estrogen-regulated gene transcription and the compromised transcriptional activity of ER-α in SNCG antisense transfected T47D cells indicated that SNCG stimulated ligand-dependent transcriptional activity of ER-α.

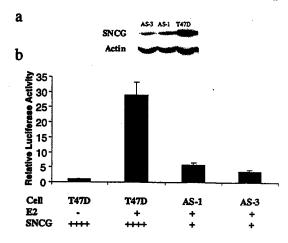
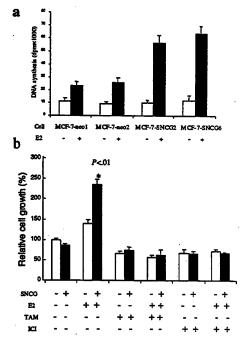


Fig. 4. Inhibition of SNCG expression reduced the transcriptional activity of ER- α . (A). Western analysis of SNCG expression in control T47D and SNCG antisense transfected AS-1 and AS-3 cells. (B). ERE-Luc reporter activity in control and antisense transfected T47D cells. Cells were cultured in the ligand-free conditioned medium for 4 days, treated with or without 10^{-11} M of E2 for 24 h before harvesting. All values were normalized to the reporter activity of the non-treated T47D cells, which was set to 1. The numbers represent means \pm SD of three cultures.

B-4. Stimulation of cell proliferation by SNCG. To determine the biological relevance of SNCG-stimulated ligand-dependent ER-α signaling, we analyzed the effect of SNCG overexpression on the growth of breast cancer cells. To determine whether SNCG overexpression affects ligand-dependent or ligand-independent cell growth, the cellular proliferation of the previously established two stable SNCG-transfected MCF-7 cell clones, MCF-SNCG2 and MCF-SNCG6, were compared with that of SNCG-negative cells, MCF-neo1 and MCF-neo2 (16). Data in Fig 5A shows that while SNCG had no significant effect on the proliferation of SNCG-transfected cells compared to MCF-neo cells in the absence of E2, overexpression of SNCG significantly stimulated the ligand-dependent proliferation. Treatment of neo clones with E2 stimulated an average cell proliferation 2.4-fold over controls. However, E2 treatment of SNCG clones resulted in an



average of 5.4-fold increase in the proliferation vs. controls, suggesting that SNCG expression renders the cells more responsive to E2-stimulated cell proliferation. To address whether the stimulatory effect of SNCG on cell growth is mediated by ER- α , we investigated the effect of the antiestrogen tamoxifen and ICI. As shown in Fig. 5B, E2-stimulated growth in both MCF-neo1 and SNCG-MCF6 cells was significantly blocked by tamoxifen and ICI. These data indicate that SNCG-stimulated cell growth is mediated by ER- α .

Fig. 5. SNCG stimulated ligand-dependent cell proliferation. For all experiments, cells were cultured and synchronized in the ligand-free Conditioned Cell Culture medium for 4 days before the hormone treatments. A, Stimulation of cell proliferation by SNCG overexpression. Cells were treated with or without 1 nM E2 for 24 hours. Cell proliferation was measured by ³H- thymidine incorporation. Data are means ± SD of three cultures. B, Effect of

antiestrogens on SNCG-stimulated cell growth. Cells were treated with or without 1 nM of E2, 1 μ M of tamoxifen, or 1 μ M of ICI for 6 days before harvesting. Media were changed every two days with fresh estrogen and antiestrogens. Cell growth was measured using a cell proliferation kit (XTT). Data are the mean \pm SD of quadruplicate cultures. Open bar represents MCF-neo1 cells; closed bar represents MCF-SNCG6 cells.

The effect of SNCG expression on cell growth was also demonstrated in the SNCG antisense construct-transfected T47D cells. SNCG antisense mRNA expression reduced SNCG protein expression to the level of 25% of that in control T47D cells (Fig. 4A). Soft agar colony assays demonstrated that the anchorage-independent growth of T47D cells expressing SNCG antisense mRNA was significantly suppressed. When cells were cultured in soft agar without E2, there were very few colonies formed in both T47D group and T47D-SNCG antisense group. Treatment of T47D cells with E2 resulted in a 19-fold increase of colonies over the non-treated cells. However, treatment of T47D cells expressing SNCG antisense mRNA with E2 resulted in only 3-fold increase over the non-treated cells (Fig. 6). These data demonstrated that inhibition of endogenous SNCG expression dramatically diminished the cell growth in response to estrogen. Consistent with its stimulatory effect on ligand-dependent cell proliferation, overexpression of SNCG did not affect the proliferation of ER-α-negative MDA-MB-435 cells (9).

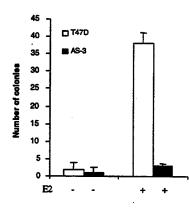


Fig. 6. Effect of inhibiting endogenous SNCG expression on soft agar colonies formation capability of T47D cells. T47D and SNCG antisense stably transfected AS-3 clone were cultured into the top layer soft agar and treated with or without 1 nM of E2 as described in Materials and Methods. The number of colonies was counted after 2 weeks of plating using a Nikon microscope at 100× amplification. Triplicate wells were assayed for each condition.

C. KEY RESEARCH ACCOMPLISHMENTS (Cancer Res., in press)

A notable finding in this study is that SNCG, previously identified as a breast cancer specific gene 1 (BCSG1), strongly stimulated the ligand-dependent transcriptional activity of estrogen receptor- α (ER- α) in breast cancer cells. Augmentation of SNCG expression stimulated transcriptional activity of ER- α , whereas compromising endogenous SNCG expression suppressed ER- α signaling. The SNCG-stimulated ER- α signaling was demonstrated in three different cell systems including ER- α -positive and SNCG-negative MCF-7 cells, ER- α -positive and SNCG-positive T47D cells, and SNCG-negative and ER- α -negative MDA-MB-435 cells. The SNCG-mediated stimulation of ER- α transcriptional activity is consistent with its stimulation of the ligand-dependent cell growth. While overexpression of SNCG stimulated the ligand-dependent cell proliferation, suppression of endogenous SNCG expression significantly inhibited cell growth in response to estrogen. The stimulatory effect of SNCG on ER α -regulated gene expression and cell growth can be effectively inhibited by antiestrogens.

D. CONCLUSIONS

1. Although synucleins are highly expressed in neuronal cells and are abundant in presynaptic terminals, synucleins have also been implicated in non-neural diseases particularly in the hormone-responsive cancers of breast and ovary. SNCG was first identified by differential cDNA sequencing as a breast cancer specific gene, which was expressed abundantly in metastatic breast cancer cDNA library but scarcely in normal breast cDNA library. SNCG expression is highly associated breast cancer and ovarian cancer progression. In addition, overexpression of SNCG in breast cancer cells significantly stimulated cell growth in vitro and tumor metastasis in vivo. However, the molecular targets of SNCG aberrant expression for breast cancer have not been identified. Here we demonstrated ER-α as one of the critical

- target molecules for SNCG's action in breast cancer pathogenesis. Thus, aberrant expression of SNCG stimulates breast cancer growth and progression by enhancing the transcriptional activity of $ER-\alpha$.
- 2. Synucleins are emerging as a central player in the fundamental neural processes and in the formation of pathologically insoluble deposits characteristic of Alzheimer's (AD) and Parkinson's (PD) diseases. Most studies of this group of proteins have been directed to the elucidation of their role in the formation of depositions in brain tissue. However, the normal cellular function of this highly conserved synuclein family remains largely unknown. Here we demonstrated that one of the functions of SNCG is activating ER-α signaling. The preventive effect of estrogen on AD has become clear with epidemiological data, suggesting that estrogen may act as a neuroprotectant against the neurodegenerative diseases. The demonstration of ER-α as one of the critical target for SNCG-mediated chaperone activity may indicate a new direction of normal cellular function of synucleins. In this regard, SNCG may be involved in mediating the function of transcriptional activity of ER-α in neuronal cells, thus, the loss or decreased SNCG expression may lower the beneficial effects of estrogen to protect neurons against PD and AD. The potential role of SNCG as a neuroprotectant warrants further investigation.

E. REFERENCES

- 1. Ji H, Liu YE, Jia T, Wang M, Liu J, Xiao G, Joseph BK and Shi YE. Identification of a breast cancer-specific gene, BCSG1, by direct differential complementary DNA sequencing. *Cancer Res.*, 57: 759-764, 1997.
- 2. Shi YE, Ni J, Xiao G, Liu YE, Fuchs A, Yu G, Zhang M, Li J, Aggarwal BB, Meager A and Gentz R. Antitumor activity of the novel human breast cancer growth inhibitor MRG. Cancer Res., 57 (15): 3084-3091, 1997.
- 3. Xiao G, Liu YE, Ni J, Gentz R, Sang QA and Shi YE. Suppression of breast cancer growth and metastasis by a novel serpin MEPI that is expressed specifically in the mammary myoepithelial cells. *PNAS* 96 (7): 3700-3705, 1999.
- 4. Jia T, Liu J, Liu YE and Shi YE. Stimulation of breast cancer invasion and metastasis by synuclein γ (SNCG). Cancer Res. 59: 742-747, 1999.
- 5. Lavedan, C., Leroy, E., Dehejia, A., Buchholtz, S., Dutra, A., Nussbaum, R. L., and Polymeropoulos, M. H. (1998). Identification, localization and characterization of the human gamma-synuclein gene. Hum Genet 103, 106-112.
- Ninkina, N. N., Alimova-Kost, M. V., Paterson, J. W., Delaney, L., Cohen, B. B., Imreh, S., Gnuchev, N. V., Davies, A. M., and Buchman, V. L. (1998). Organization, expression and polymorphism of the human persyn gene. Hum Mol Genet 7, 1417-1424.
- 7. Clayton, D. F., and George, J. M. (1998). The synucleins: a family of proteins involved in synaptic function, plasticity, neurodegeneration and disease. Trends Neurosci 21, 249-254.
- 8. Polymeropoulos, M. H., Lavedan, C., Leroy, E., Ide, S. E., Dehejia, A., Dutra, A., Pike, B., Root, H., Rubenstein, J., Boyer, R., et al. (1997). Mutation in the alpha-synuclein gene identified in families with Parkinson's disease. Science 276, 2045-2047.
- 9. Spillantini, M. G., Schmidt, M. L., Lee, V. M., Trojanowski, J. Q., Jakes, R., and Goedert, M. (1997). Alpha-synuclein in Lewy bodies. Nature 388, 839-840.
- 10. Takeda, A., Mallory, M., Sundsmo, M., Honer, W., Hansen, L., and Masliah, E. (1998). Abnormal accumulation of NACP/alpha-synuclein in neurodegenerative disorders. Am J Pathol 152, 367-372.
- 11. Ueda, K., Fukushima, H., Masliah, E., Xia, Y., Iwai, A., Yoshimoto, M., Otero, D. A., Kondo, J., Ihara, Y., and Saitoh, T. (1993). Molecular cloning of cDNA encoding an unrecognized component of amyloid in Alzheimer disease. Proc Natl Acad Sci U S A 90, 11282-11286.
- 12. Yoshimoto, M., Iwai, A., Kang, D., Otero, D. A., Xia, Y., and Saitoh, T. (1995). NACP, the precursor protein of the non-amyloid beta/A4 protein (A beta) component of Alzheimer disease amyloid, binds A beta and stimulates A beta aggregation. Proc Natl Acad Sci U S A 92, 9141-9145.
- 13. Duda, J. E., Lee, V. M., and Trojanowski, J. Q. (2000). Neuropathology of synuclein aggregates. J Neurosci Res 61, 121-127.

- 14. Galvin, J. E., Uryu, K., Lee, V. M., and Trojanowski, J. Q. (1999). Axon pathology in Parkinson's disease and Lewy body dementia hippocampus contains alpha-, beta-, and gamma-synuclein. Proc Natl Acad Sci U S A 96, 13450-13455.
- 15. Liu A, Gupta A, Li C, Ahlborn TE, Ma Y, Shi YE and Liu J. Molecular Mechanisms for Aberrant Expression of the Human Breast Cancer Specific Gene 1 in Breast Cancer Cells: Control of transcription by DNA methylation and intronic sequences. Oncogene 20 (37): 5173-5185, 2001.
- 16. Liu J, Spence MJ, Zhang YL, Jiang Y, Liu YE and Shi YE. Transcriptional suppression of synuclein γ (SNCG) expression in human breast cancer cells by the growth inhibitory cytokine oncostatin M. Breast Cancer Research and Treatment 62:.99-107, 2000.
- 17. Bruening, W., Giasson, B. I., Klein-Szanto, A. J., Lee, V. M., Trojanowski, J. Q., and Godwin, A. K. (2000). Synucleins are expressed in the majority of breast and ovarian carcinomas and in preneoplastic lesions of the ovary. Cancer 88, 2154-2163.
- 18. Perlmann, T., and Evans, R.M. Nuclear receptors in Sicily: all in the famiglia. Cell, 90: 391-397, 1997.
- 19. Katzenellenbogen, B.S. Estrogen receptors: bioactivities and interactions with cell signaling pathways. Biol. Reprod., 54: 287-293, 1996.